

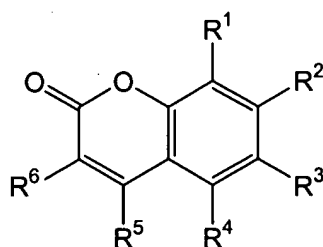
**Amendments to the Claims:**

This listing of claims will replace all prior versions, and listings of claims in the application:

**Listing of Claims:**

1-83 (canceled)

84 (new): A material having a fluorogenic moiety linked to a solid support, said material having the structure:



wherein:

R<sup>1</sup>, R<sup>3</sup>, R<sup>4</sup> and R<sup>6</sup> are each H;

R<sup>2</sup> is -NHR<sup>15</sup>; and

R<sup>5</sup> is -R<sup>14</sup>-SS,

wherein:

R<sup>14</sup> is -CH<sub>2</sub>C(O)NH-;

R<sup>15</sup> is a member selected from the group consisting of amine protecting groups, -C(O)-AA and -C(O)-P:

wherein:

P is a peptide sequence;

AA is an amino acid residue; and

SS is a solid support.

1                   **85** (new): The material in accordance with claim **84**, wherein  $R^{15}$  is an amine  
2 protecting group.

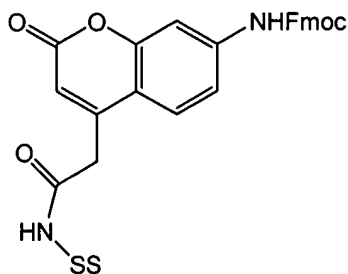
1                   **86** (new): The material in accordance with claim **85**, wherein said amine  
2 protecting group is 9-fluorenylmethoxycarbonyl (Fmoc).

1                   **87** (new): The material in accordance with claim **84**, wherein  $R^{15}$  is  $-C(O)-AA$ ,  
2 wherein AA is an amino acid residue.

1                   **88** (new): The material in accordance with claim **84**, wherein  $R^{15}$  is  $-C(O)-P$ ,  
2 wherein P is a peptide sequence.

1                   **89** (new): The material in accordance with claim **84**, wherein the solid support is  
2 a Rink resin.

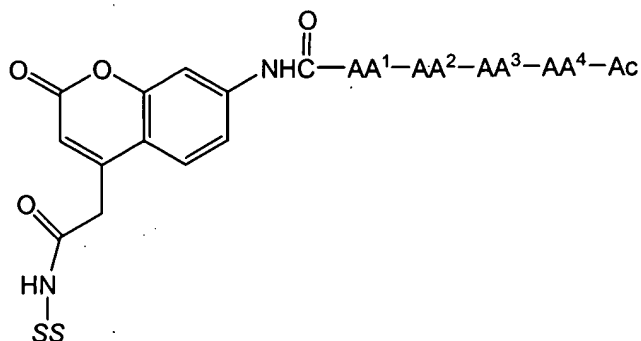
1                   **90** (new): A material having a fluorogenic moiety linked to a solid support, said  
2 material having the structure:



3  
wherein:

4                   SS is a solid support, wherein said the support is a Rink resin.

1                   **91** (new): A library of fluorogenic peptides comprising sub-libraries P1, P2, P3  
2 and P4, wherein each of the sub-libraries P1, P2, P3 and P4 comprises tetrapeptides having the  
3 structure:



wherein:

SS is a solid support, and

wherein:

for sub-library P1, each AA¹ is a different amino acid of the 20 amino acids, and each of AA²-AA⁴ is an isokinetic mixture of 20 amino acids;

for sub-library P2, each of AA² is a different amino acid of the 20 amino acids, and each of AA¹, AA³ and AA⁴ is an isokinetic mixture of 20 amino acids;

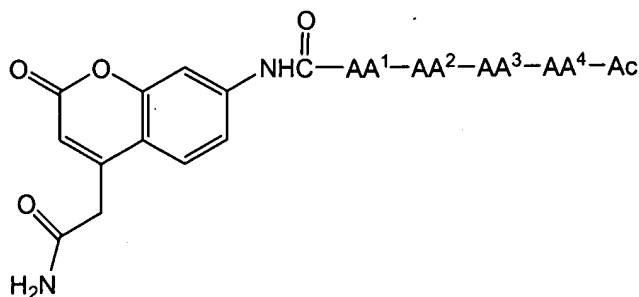
for sub-library P3, each of AA³ is a different amino acid of the 20 amino acids, and each of AA¹, AA² and AA⁴ is an isokinetic mixture of 20 amino acids; and

for sub-library P4, each of AA⁴ is a different amino acid of the 20 amino acids, and each of AA¹, AA² and AA³ is an isokinetic mixture of 20 amino acids.

**92 (new):** The library in accordance with claim 91, wherein the 20 amino acids are the 20 naturally occurring amino acids excluding cysteine and including norleucine.

**93 (new):** The library in accordance with claim 91, wherein the solid support is a Rink resin.

**94 (new):** A library of fluorogenic peptides comprising sub-libraries P1, P2, P3 and P4, wherein each of the sub-libraries P1, P2, P3 and P4 comprises tetrapeptides having the structure:



wherein:

for sub-library P1, each  $\text{AA}^1$  is a different amino acid of the 20 amino acids, and each of  $\text{AA}^2$ - $\text{AA}^4$  is an isokinetic mixture of 20 amino acids;

for sub-library P2, each of  $\text{AA}^2$  is a different amino acid of the 20 amino acids, and each of  $\text{AA}^1$ ,  $\text{AA}^3$  and  $\text{AA}^4$  is an isokinetic mixture of 20 amino acids;

for sub-library P3, each of  $\text{AA}^3$  is a different amino acid of the 20 amino acids, and each of  $\text{AA}^1$ ,  $\text{AA}^2$  and  $\text{AA}^4$  is an isokinetic mixture of 20 amino acids; and

for sub-library P4, each of  $\text{AA}^4$  is a different amino acid of the 20 amino acids, and each of  $\text{AA}^1$ ,  $\text{AA}^2$  and  $\text{AA}^3$  is an isokinetic mixture of 20 amino acids.

**95 (new):** The library in accordance with claim 94, wherein the 20 amino acids are the 20 naturally occurring amino acids excluding cysteine and including norleucine.

**96 (new):** A method of determining a peptide sequence specificity profile of an enzymatically active protease, said method comprising:

(a) contacting said protease with a library of peptides according to claim 91 or claim 94 in such a manner whereby the fluorogenic moiety is released from the peptide sequence, thereby forming a fluorescent moiety;

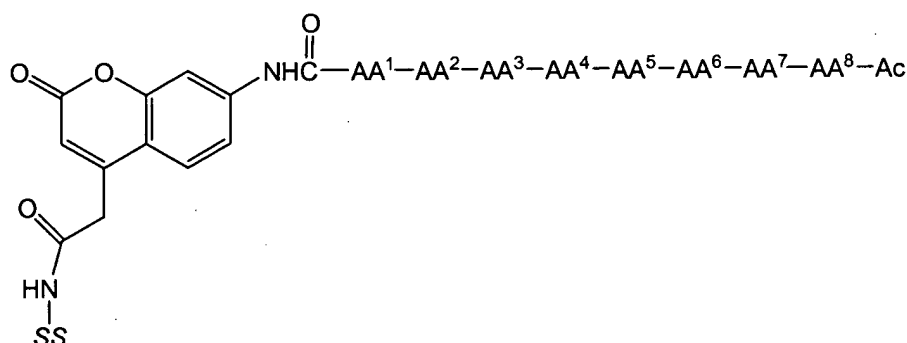
(b) detecting said fluorescent moiety;

(c) determining the sequence of said peptide sequence, thereby determining said peptide sequence specificity profile of said protease.

1                    **97 (new):** The method according to claim **96**, further comprising (d) quantifying  
2 said fluorescent moiety, thereby quantifying said protease.

1                    **98 (new):** The method according to claim **97**, wherein said protease is a member  
2 selected from the group consisting of aspartic protease, cysteine protease, metalloprotease and  
3 serine protease.

1                    **99 (new):** A library of fluorogenic peptides comprising sub-libraries P1, P2, P3  
2 and P4, wherein each of the sub-libraries P1, P2, P3 and P4 comprises hexapeptides having the  
3 structure:



4  
5 wherein:

6                    SS is a solid support, and

7 wherein:

8                    for each sub-library P1, P2, P3 and P4, AA¹, AA², AA³ and AA⁴ in each of the  
9 hexapeptides are the same amino acid residues;

10                    for sub-library P1, each of AA⁵ is a different amino acid of the 20 amino acids,  
11 and each of AA⁶, AA⁷ and AA⁸ is an isokinetic mixture of 20 amino acids;

12                    for sub-library P2, each of AA⁶ is a different amino acid of the 20 amino acids,  
13 and each of AA⁵, AA⁷ and AA⁸ is an isokinetic mixture of 20 amino acids;

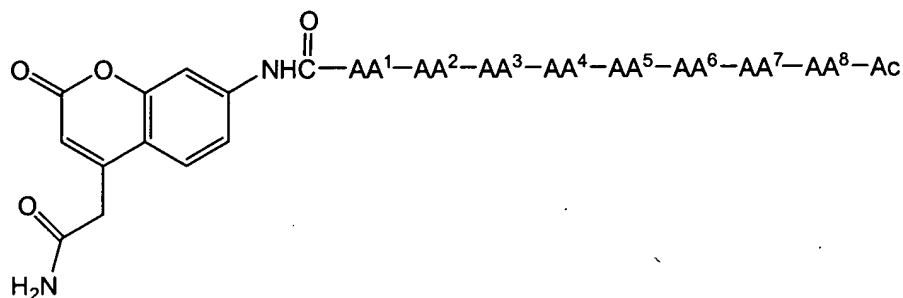
14                    for sub-library P3, each of AA⁷ is a different amino acid of the 20 amino acids,  
15 and each of AA⁵, AA⁶ and AA⁸ is an isokinetic mixture of 20 amino acids; and

for sub-library P4, each of AA<sup>8</sup> is a different amino acid of the 20 amino acids,  
and each of AA<sup>5</sup>, AA<sup>6</sup> and AA<sup>7</sup> is an isokinetic mixture of 20 amino acids.

**100 (new):** The library in accordance with claim **99**, wherein the 20 amino acids  
are the 20 naturally occurring amino acids excluding cysteine and including norleucine.

**101 (new):** The library in accordance with claim **99**, wherein the solid support is  
a Rink resin.

**102 (new):** A library of fluorogenic peptides comprising sub-libraries P1, P2, P3  
and P4, wherein each of the sub-libraries P1, P2, P3 and P4 comprises hexapeptides having the  
structure:



wherein:

for each sub-library P1, P2, P3 and P4, AA<sup>1</sup>, AA<sup>2</sup>, AA<sup>3</sup> and AA<sup>4</sup> in each of the  
hexapeptides are the same amino acid residues;

for sub-library P1, each of AA<sup>5</sup> is a different amino acid of the 20 amino acids,  
and each of AA<sup>6</sup>, AA<sup>7</sup> and AA<sup>8</sup> is an isokinetic mixture of 20 amino acids;

for sub-library P2, each of AA<sup>6</sup> is a different amino acid of the 20 amino acids,  
and each of AA<sup>5</sup>, AA<sup>7</sup> and AA<sup>8</sup> is an isokinetic mixture of 20 amino acids;

for sub-library P3, each of AA<sup>7</sup> is a different amino acid of the 20 amino acids,  
and each of AA<sup>5</sup>, AA<sup>6</sup> and AA<sup>8</sup> is an isokinetic mixture of 20 amino acids; and

for sub-library P4, each of AA<sup>8</sup> is a different amino acid of the 20 amino acids,  
and each of AA<sup>5</sup>, AA<sup>6</sup> and AA<sup>7</sup> is an isokinetic mixture of 20 amino acids.

1                   **103** (new): The library in accordance with claim **102**, wherein the 20 amino  
2 acids are the 20 naturally occurring amino acids excluding cysteine and including norleucine.

1                   **104** (new): A method of determining a peptide sequence specificity profile of an  
2 enzymatically active protease, said method comprising:

3                   (a) contacting said protease with a library of peptides according to claim **99** or  
4                   claim **102** in such a manner whereby the fluorogenic moiety is released  
5                   from the peptide sequence, thereby forming a fluorescent moiety;

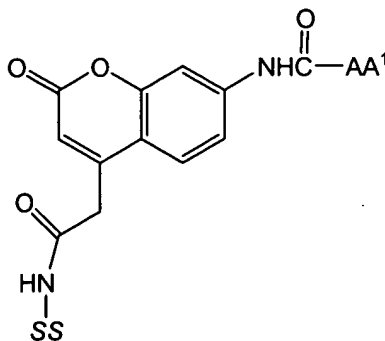
6                   (b) detecting said fluorescent moiety;

7                   (c) determining the sequence of said peptide sequence, thereby determining said  
8                   peptide sequence specificity profile of said protease.

1                   **105** (new): The method according to claim **104**, further comprising (d)  
2 quantifying said fluorescent moiety, thereby quantifying said protease.

1                   **106** (new): The method according to claim **105**, wherein said protease is a  
2 member selected from the group consisting of aspartic protease, cysteine protease,  
3 metalloprotease and serine protease.

1                   **107** (new): A library of twenty fluorogenic amino acid amides having the  
2 structure:



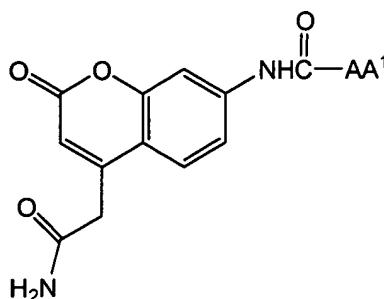
3  
4                   wherein:

5                    *SS* is a solid support, and  
6                    each AA<sup>1</sup> for the twenty fluorogenic amino acid amides is a different amino acid  
7 residue.

1                    **108** (new): The library in accordance with claim **107**, wherein the amino acid  
2 residues are the 20 naturally occurring amino acids excluding cysteine and including norleucine.

1                    **109** (new): The library in accordance with claim **108**, wherein the solid support  
2 is a Rink resin.

1                    **110** (new): A library of twenty fluorogenic amino acids having the structure:



2  
3 wherein:

4                    each AA<sup>1</sup> for the twenty fluorogenic amino acids is a different amino acid residue

1                    **111** (new): The library in accordance with claim **110**, wherein the amino acid  
2 residues are the 20 naturally occurring amino acids excluding cysteine and including norleucine..

1                    **112** (new): A method of determining an amino acid specificity profile of an  
2 enzymatically active protease, said method comprising:

3                    (a) contacting said protease with a library of amino acids according to claim **108**  
4    or claim **110** in such a manner whereby the fluorogenic moiety is released  
5    from the amino acid, thereby forming a fluorescent moiety;

6                    (b) detecting said fluorescent moiety;



7                   (c) determining the identity of the amino acid, thereby determining said amino  
8                   acid specificity profile of said protease.

1                   **113 (new):** The method according to claim **112**, further comprising (d)  
2                   quantifying said fluorescent moiety, thereby quantifying said protease.

1                   **114 (new):** The method according to claim **113**, wherein said protease is a  
2                   member selected from the group consisting of aspartic protease, cysteine protease,  
3                   metalloprotease and serine protease.